Fluorescence Analysis Activity

1 Calculating Fluorescence Area

- 1. Open up ImageJ and open demo2 1.png from the BMES Cell Team website
- 2. Click on Image > Adjust > Color threshold to separate the cells from the background
- 3. Go to Analyze > Analyze Particles > Display results
- 4. Highlight the entire table, click on File > Save as, and save your file as a comma separated value (csv).

Feel free to name your file whatever you want (tacos.csv, screwthis.csv, etc.)

- 5. Open up Anaconda and click on Spyder
- 6. Follow along with the demo

2 Calculating the Fluorescence Intensity

- 1. Close ImageJ and reopen it again
- 2. Open demo2 1.png from the BMES Cell Team website
- 3. Click on Image > Type > 8-bit
- 4. Click on Analyze > Set Measurements. Make sure Area, Integrated Density, and Mean Grey Value are checked.
- 5. Under the toolbar, click on the freehand tool
- 6. With your mouse, trace around the outline of a particular cell
- 7. Click on Analyze > Measure. If you have a Mac, you can use Command + M. A window will pop up.
- 8. Repeat steps 5-7 three more times for different cells. You should have four different measurements.
- 9. Copy the information to your spreadsheet.
- 10. Now, find a small area of your image that has no fluorescence. This will be your background.
- 11. Click on the freehand tool and trace around this small area.
- 12. Click on Analyze > Measure (or Command + M if you have a Mac).
- 13. Repeat steps 10-12 three more times for different background areas. You should have a total of 4 background measurements.
- 14. Copy the information to your spreadsheet.

Your final spreadsheet should look like this, but with different values for Area, Mean, and Integrated Density:

Cell	Area	Mean	Integrated Density
1	385	39.681	15277
2	494	26.097	12892
3	226	41.615	9405
4	406	40.631	16496
Background			
1	277	0.152	42
2	183	0	0
3	72	0	0
4	74	0.243	18

^{15.} Open up Anaconda and click on Spyder

^{16.} Follow along with the demo